

## Alternative methods of semen preservation: A pig model for endangered species

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#### Contents





# Materials and Methods Results





# A high valuable animal suddenly died and had no preserved semen !



# "Please, help me doctor!" "How can I get a descendant from my dead animal ?"





There are less than 250 wild Riverine rabbits left in South Africa

Bunolagus monticularis; Red list (IUCN): critically endangered species

http://www.edgeofexistence.org/mammals/species\_info.php?id=3





#### Tuaklom † 25.06.2010



Some methods for desiccated semen preservation

Freeze-drying

(Bhowmick et al. 2003, McGinnis et al. 2005)

Air-drying (Imoedamhe 2005) مر

Requirement: drying chamber, gas releasing

regulator, electric fan and larminar flow chamber



# Some methods for desiccated semen preservation

Heat-drying (Lee and Niwa 2006, Rungroekrit et al. 2012, Lee et al. 2013)

Flame-drying (Rungroekrit et al. 2013)



Simple semen preservation method

Less equipment requirement

- Portable stuffs
  - Preserved sperm can be stored at
    - ambient temperature for short- and/or

long- term

#### Goal



#### **Heat- and Flame-drying** as an alternative

methods (simple procedure and possible to use under extreme condition area)

Fertilization ability investigation using ICSI

Investigation of sperm DNA fragmentation using Halomax<sup>®</sup> test kit



#### In vitro maturation

- ≻Cumulus oocyte complexes (COCs) ≥ 3 layers
- IVM in mTCM 199 (insulin, L-glutamine, FCS, gentamicin; stock medium) and eCG
- Maturation period: 44 46 h (39 °C, 5 % CO<sub>2</sub>, humidified atmosphere)





Sperm preparation: Heat-drying





#### Sperm preparation: Flame-drying





Dried sperm samples packaging and storage





Dried sperm sample rehydration





#### Sperm DNA fragmentation assessment





#### Fertilization ability testing using ICSI



#### Micromanipulation unit



#### Intracytoplasmic sperm injection

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Sperm-injected oocytes activation and *in vitro* culture

10 % ethanol (2 min) in mTCM 199

/ mTCM 199 + 10 µg/ml CHX (24 h)

staining with aceto-orcein









# Sperm DNA fragmentation by Halomax<sup>®</sup> test kit



#### Results



#### Percent of DNA fragmentation index (% DFI)

Spermatozoa	Storage	No. of	Total no. of examined	% DFI
	duration	replicates	spermatozoa	(mean± SD)
Ejaculated semen (control gr.)		4	1,200	$1.5 \pm 0.4^{a}$
HD 50 °C 45 min	short-term	4	1,200	59.66 ± 2.0°
	long-term	4	1,200	$68.66 \pm 1.9^{c^*}$
HD 56 °C 45 min	short-term	4	1,200	58.17 ± 3.8°
	long-term	4	1,200	64.58 ± 3.1 <sup>c*</sup>
HD 90 °C 45 min	short-term	4	1,200	$75.0 \pm 4.0^{d}$
	long-term	4	1,200	$82.66 \pm 2.8^{d}$
HD 120 °C 20 min	short-term	4	1,200	<b>1.25 ± 0.7</b> <sup>a</sup>
	long-term	4	1,200	<b>2.33 ± 0.2</b> <sup>a*</sup>
FF	short-term	4	1,200	$38.0 \pm 4.0^{E}$
	long-term	4	1,200	$50.58 \pm 4.3^{e^*}$
FSW	short-term	4	1,200	$30.25 \pm 3.5^{F}$
	long-term	4	1,200	$34.0 \pm 3.2^{f}$

HD: Heat-dried semen samples, FF: Flame-dried sperm rich fraction, FSW: Flame-dried swim up semen, *P*<0.05

## Results



#### Fertilization criteria



Two polar bodies, formation of 2 pronuclei, sperm tail and disappearence of sperm head, bar =  $20 \ \mu m$ 

#### Results



#### Percent of fertilized oocytes after ICSI

Experimental group	Storage duration	No. of examined oocytes	No. (%) of fertilized oocytes
Ejaculated semen (control gr.)		140	26 (18.6)
HD 50 °C 45 min	short-term	140	20 (14.3)
	long-term	124	7 (5.7)
HD 56 °C 45 min	short-term	122	21 (17.2)
	long-term	103	9 (8.7)
HD 90 °C 45 min	short-term	135	15 (11.1)
	long-term	142	22 (15.5)
HD 120 °C 20 min	short-term	102	0 (0)
	long-term	122	0 (0)
<b>FF</b>	short-term	128	14 (10.9)
rr	long-term	126	10 (7.9)
FSW	short-term	115	12 (10.4)
	long-term	130	15 (11.5)

HD: Heat-dried semen samples, FF: Flame-dried sperm rich fraction, FSW: Flame-dried swim up semen, 5-7 replications per sample

#### Conclusions



Heat- and Flame-dried boar semen have an ability to fertilize porcine oocytes by using ICSI technique

Long-term storage increased % DFI of dried samples

Further experiments need to optimize the protocols in term of sperm DNA damage protection, storage method, and rehydration procedure to improve the fertilization rate



# Thank you for your attention

